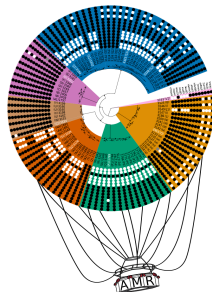


Around the resistome in 80 ways:

an empirical evaluation of antimicrobial
resistance gene detection methods



Finlay Maguire

finlaymaguire@gmail.com

December 2, 2019

Faculty of Computer Science, Dalhousie University

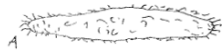
Table of contents

1. Background
2. Why do we care about AMR?
3. Targeted sequencing
4. Genomics
5. Metagenomics
6. Metagenomic-Assembled Genomes

Background

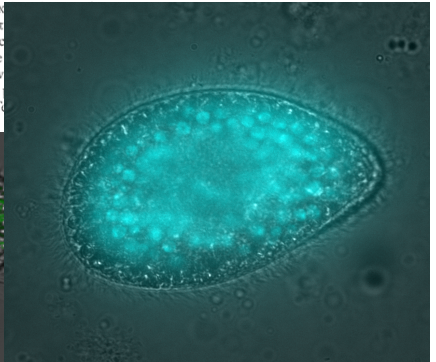
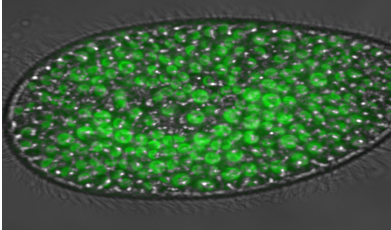
Evolution of Eukaryotic Endosymbioses

peu. Et par deux fois j'ay vu dans cette mefine eau un animal dix
que ces autres qui avoit des pieds tout le long du corps, et estoit

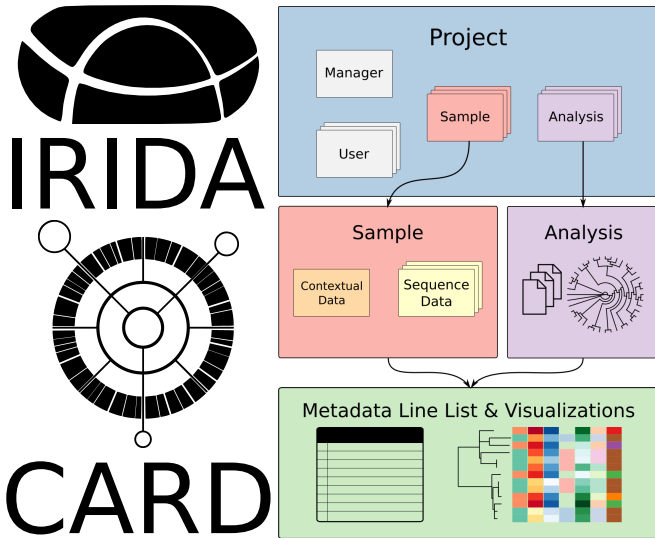


Les 4 ou 5 pieds du c
fans celle quand mefine
en repos. Il courroit v
autres, et se tournoit et
l'eau. Hartfoecker m'aff

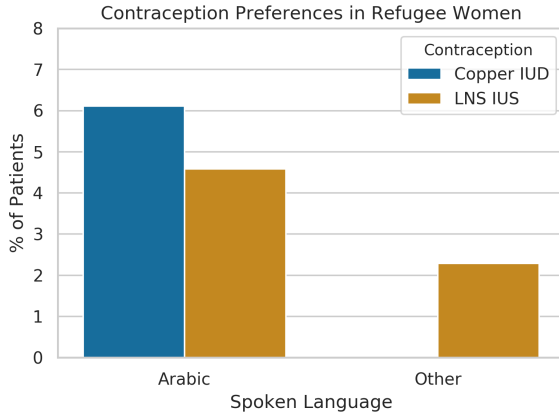
trouvè de la mefine espee in femine corrupto.



(Maguire, 2016)

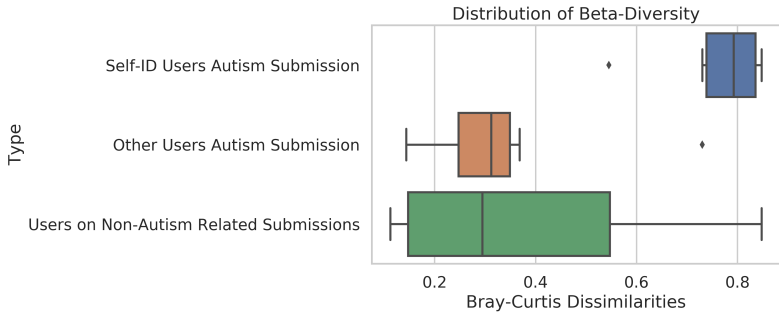


(Matthews et al., 2018)



(Stairs et al., 2019)

Sociology?

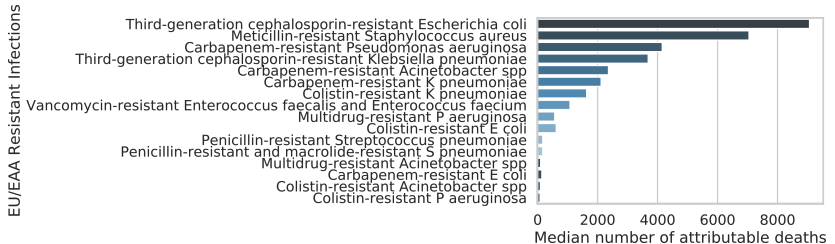


Congratulations, your application to the SSHRC Explore Grants competition has been awarded.

Project Title: NEETs, Incels, and Wizards: The Experiences of Socially Isolated Men

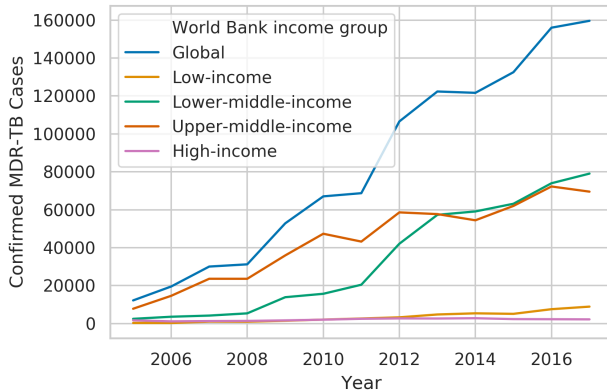
Why do we care about AMR?

AMR is currently a problem



2015 EU/EEA: 33,110 deaths, Data from (Cassini et al., 2019).

AMR is growing



WHO Global Health Observatory Data Repository.

What can we do about it?

Improve surveillance

- Locally: information would help improve patient health.

Improve surveillance

- Locally: information would help improve patient health.
- Nationally: health policies and responses to emergencies.

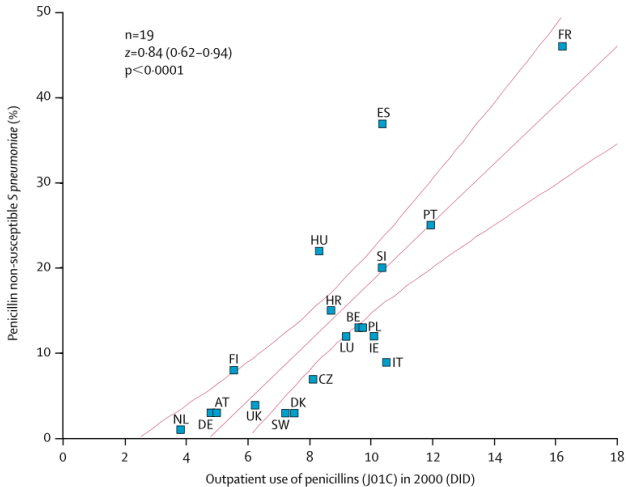
Improve surveillance

- Locally: information would help improve patient health.
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- Globally: emerging threats and long-term trends.

Improve surveillance

- Locally: information would help improve patient health.
- Nationally: health policies and responses to emergencies.
- Globally: emerging threats and long-term trends.
- Scientifically: better understanding of underlying biology.

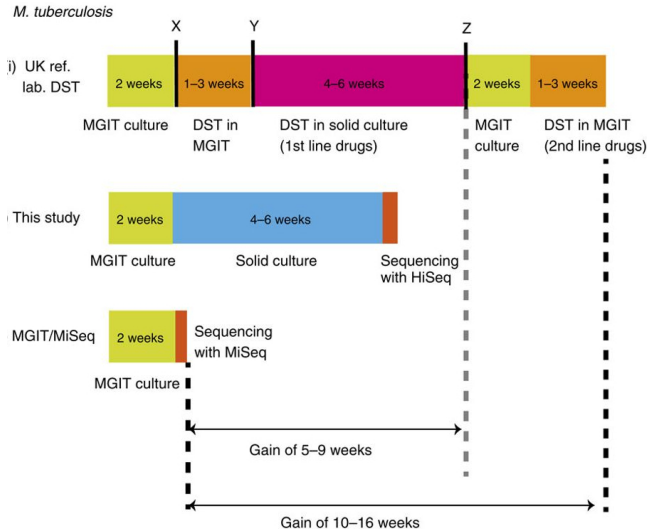
Improve diagnostics



(Goossens et al., 2005)

How do we do this?

Phenotypically?



(Bradley et al., 2015)

DNA sequencing

- DNA is relatively tractable and stable
- Sequencing technology is mature
- Represents the substrate of evolution

***E. coli* gene regulatory networks are inconsistent with gene expression data**

Simon J Larsen , Richard Röttger, Harald H H W Schmidt, Jan Baumbach

Nucleic Acids Research, Volume 47, Issue 1, 10 January 2019, Pages 85–92,

Random sequences rapidly evolve into de novo promoters

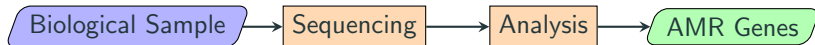
Avihu H. Yona , Eric J. Alm & Jeff Gore 

Nature Communications **9**, Article number: 1530 (2018) | [Cite this article](#)

- 10% of random sequences can serve as active promoters
- 60% of random sequences can modulate expression with only one mutation

Which DNA sequencing method?

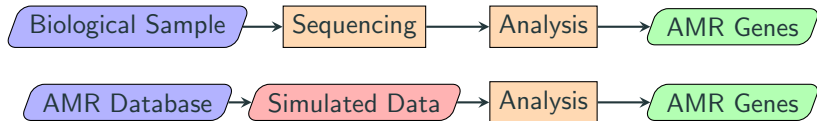
Choosing a method



Additional factors:

- Does method provide other information?
- Cost/experimental considerations

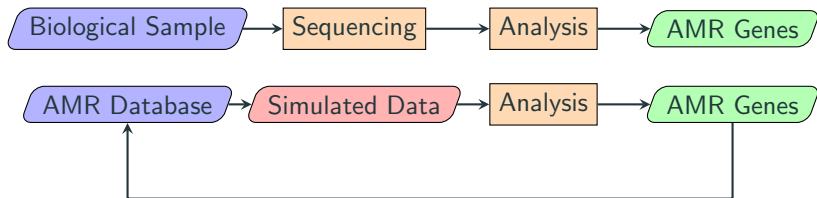
Choosing a method



Additional factors:

- Does method provide other information?
- Cost/experimental considerations

Choosing a method

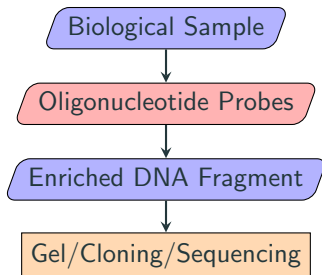


Additional factors:

- Does method provide other information?
- Cost/experimental considerations

Targeted sequencing

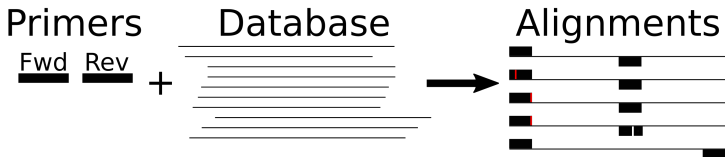
Targeted sequencing



- Cheap/simple infrastructure
- Multiple sample types
- Low input requirements

Choosing and evaluating primers

Testing primers computationally



github.com/mwhall/VAware: Mike Hall

Needleman-Wunsch alignments:

- Perfect: no mismatches, insert < 1500
- Intermediate: (1-2 minor mismatches)
- Low: (2-4 minor; 0-1 major - terminal, gaps)
- Missed: (> 4 minor; > 1 major)

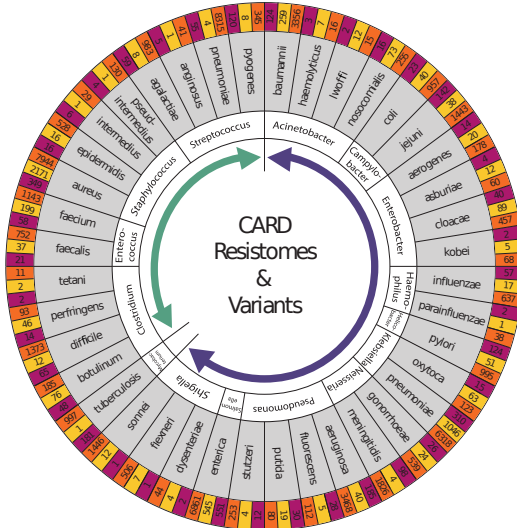
Which primers?

List of primers for detection of antimicrobial resistance genes

Antimicrobial	Genus	Gene/locus no.	Primer name	Internal number	Sequence	Temperature (°C)	Reference
Beta-lactams	TEM	All	TEM front P1	Primer 757	5'-GGCGAACCCCTATTGG-3'	55	Olsson, I. H. Holmström, and F. M. Aarestrup. 2004. Prevalence of beta-lactamases among ampicillin-resistant <i>Escherichia coli</i> and <i>Salmonella</i> isolated from food animals in Denmark. <i>Microb Drug Resist</i> 10:134-145.
			TEM-C-R-ep	Primer 686	5'-ATC AAT GCT TAA TCA GTG AG-3'		
	CTX	M-AB	ctx-M U1	Primer 1354	5'-ATGTGCAGVACCAATAATGATGCG-3'	60	Hartman, H., D. Mevius, K. Volden, J. Olsson and P. M. Aarestrup. 2006. Beta-lactamases among Extended spectrum Beta-lactamase resistant (ESBL) <i>Salmonella</i> from poultry, poultry products and human patients in The Netherlands. <i>J. Antimicrob. Chemother.</i> 56:115-121.
			CTX-M-U-2new	Primer 1580	5'-TAGGTAAATATGATGACAGAAVAGCGCG-3'		
		CTX-M1 group	ctx-M-15 front P1	Primer 1537	5'-CCATGGTAAAAAATCACTGCG-3'		
							Hendriksen, R. S., Munkittrick, K., Kirschebaum, C., Rickert, R. L., Daye, S. V., Khatib, C., Hansen, H., Corcoran, M., Mevius, D., Threlkeld, J., Angulo, P. J., Aarestrup, P. M. 2009. Emergence of Multidrug-Resistant <i>Salmonella</i> Concord Infections in Europe and the United States in Children Adopted From Ethiopia, 2005-2007. <i>Pediatr Infect Dis J</i> 3: 29:814-818.
							Moodley, A. and Garudubawa, I. Transmission of IncN Plasmids Carrying blaCTX-M-1 between Commensal <i>Escherichia coli</i> in Pigs and Farm Workers. <i>Antimicrobial Agents and Chemotherapy</i> . 2009. 53:1709-1711.

European Committee on Antimicrobial Susceptibility Testing: 78 PCR Primer Sets

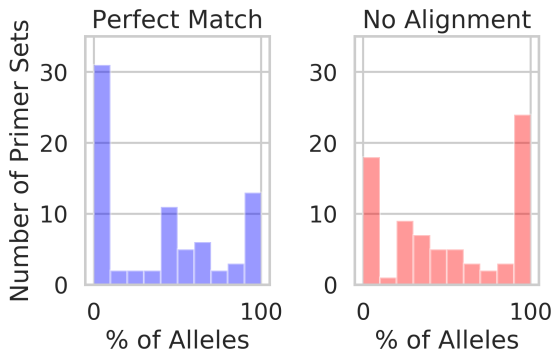
Which AMR genes?



CARD-prevalence: 85 pathogens, 116,914 resistomes (chromosome, plasmid, and WGS assembly). Brian Alcock/McArthur Lab

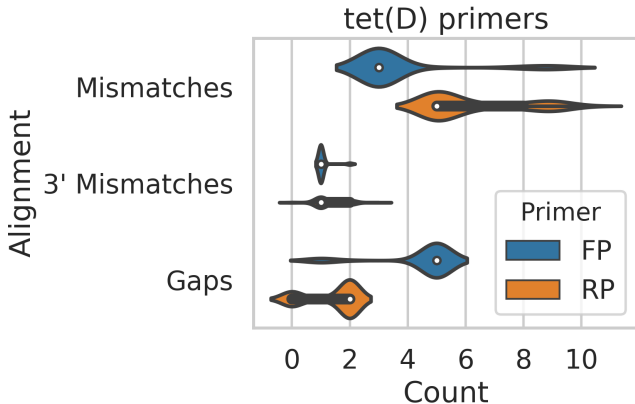
How well do these primers work?

Surprisingly poorly



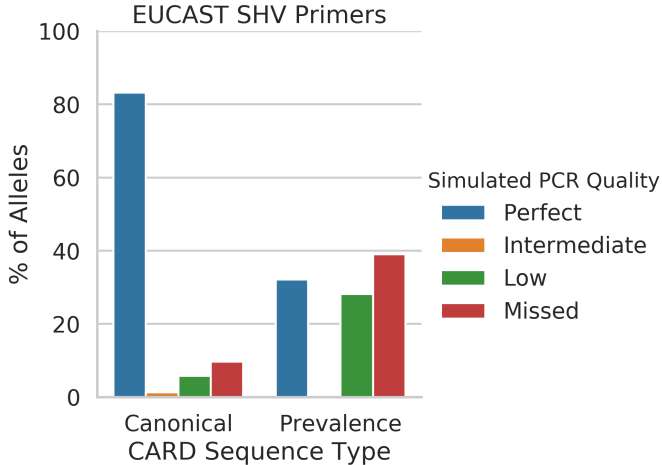
- Many aminoglycosides and tetracycline resistance genes totally missed
- Caveat: needs experimental validation

Lots of serious mismatches



No primer alignment in 27.58% of *tetD* alleles

Stagnation of primers



off-target hits (1 mismatch in RP) to *LEN-3*, *LEN-4*

Can we improve on this?



AMERICAN
SOCIETY FOR
MICROBIOLOGY

Antimicrobial Agents
and Chemotherapy

Mechanisms of Resistance

Capturing the Resistome: A targeted capture method to reveal antibiotic resistance determinants in metagenomes

Allison K. Guiton, Amogelang R. Raphenya, Jennifer Klunk, Melanie Kuch, Brian Alcock, Michael G. Surette, Andrew G. McArthur, Hendrik N. Poinar, Gerard D. Wright

DOI: 10.1128/AAC.01324-19

(Guiton et al., 2019)

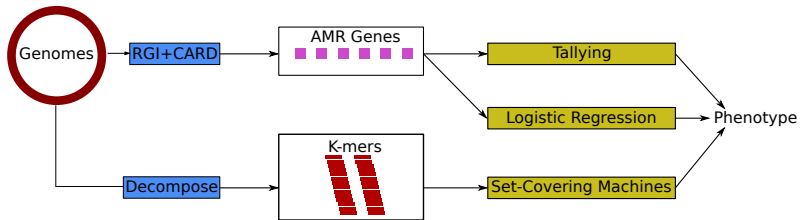
Downsides of targeted-approaches

- *a priori* target decisions
- Need constantly updated
- No easy genomic context
- No easy source-genome attribution

Why do we care about context?

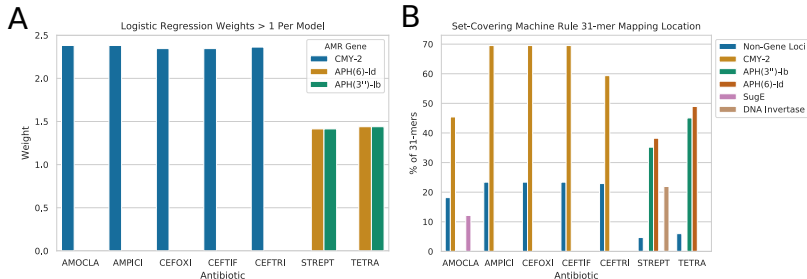
Genomics

Phenotype prediction modelling



(Maguire et al., 2019)

Genomes allow gene-free models



(Maguire et al., 2019)

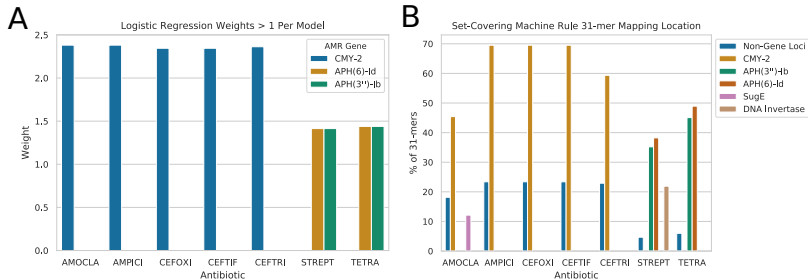
Appl Environ Microbiol, 2011 Jul;77(13):4486-93. doi: 10.1128/AEM.02788-10. Epub 2011 May 20.

Selection pressure required for long-term persistence of blaCMY-2-positive IncA/C plasmids.

Subbiah M¹, Top EM, Shah DH, Call DR.

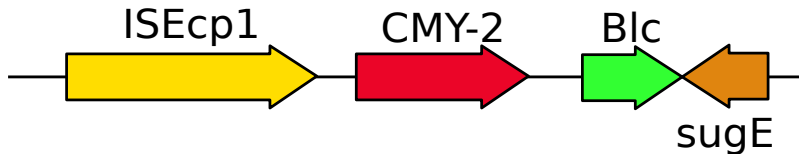
(Maguire et al., 2019)

Generate co-selection hypotheses



(Maguire et al., 2019)

Generate co-selection hypotheses



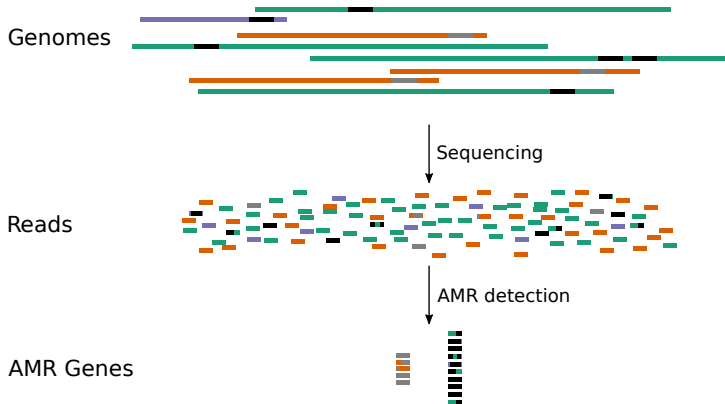
(Maguire et al., 2019)

We need genomes to identify previously unknown factors, but:

- Culturing is expensive, time-consuming, and difficult
- Single cell methods are noisy and analytically complex
- Only profile 'one' genome per sample

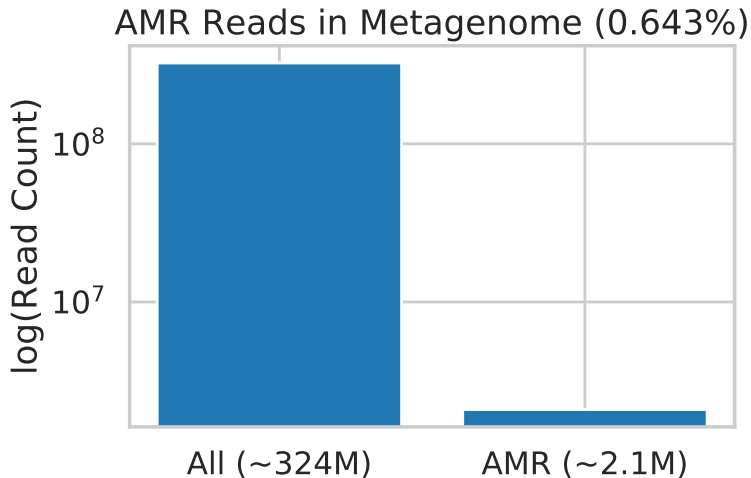
Metagenomics

Read-based AMR Metagenomics



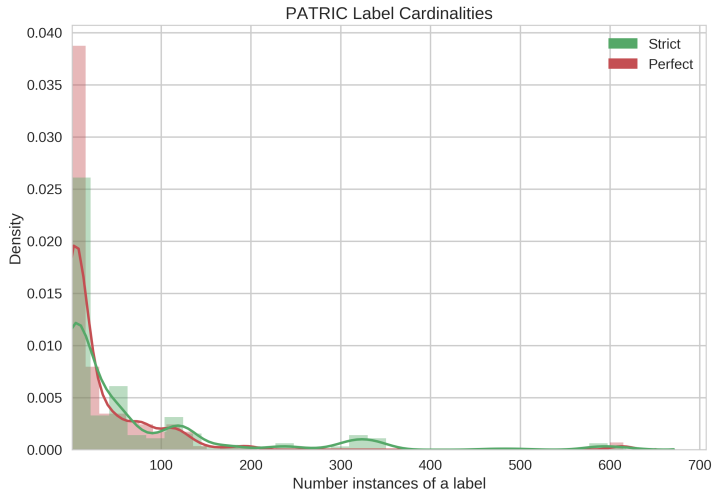
Difficulties of metagenomics

AMR genes are rare genomically



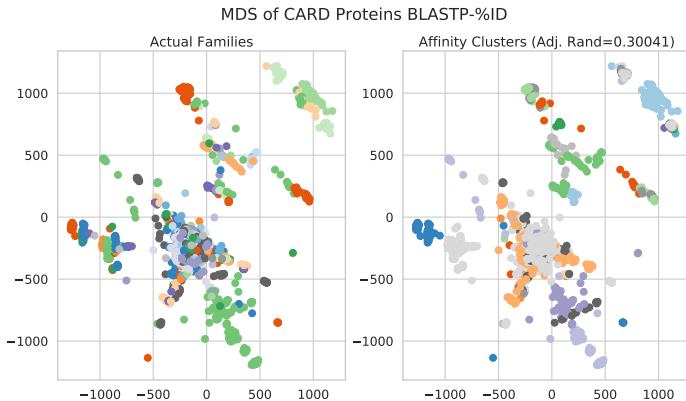
2184 CARD-prevalence genomes at 1-10X abundance

AMR genes have wildly different abundances



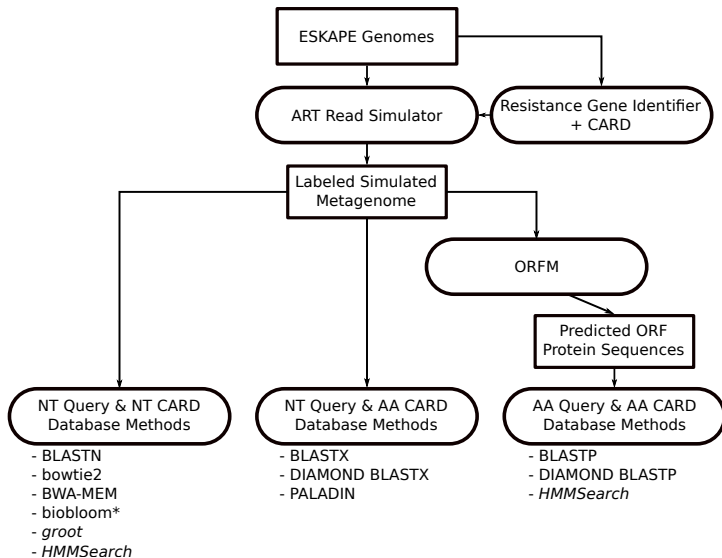
1236 AMR PATRIC genomes

AMR sequence space overlaps

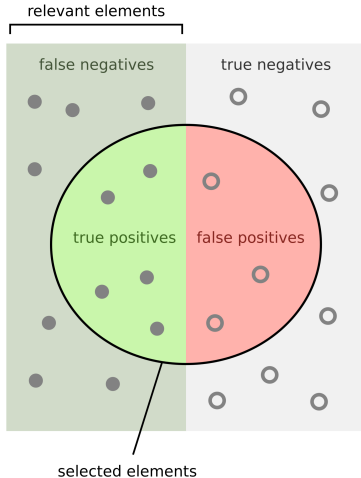


Choosing an analysis approach

Simulate data and compare tools



Terminology refresher



How many selected items are relevant?

$$\text{Precision} = \frac{\text{true positives}}{\text{true positives} + \text{false positives}}$$

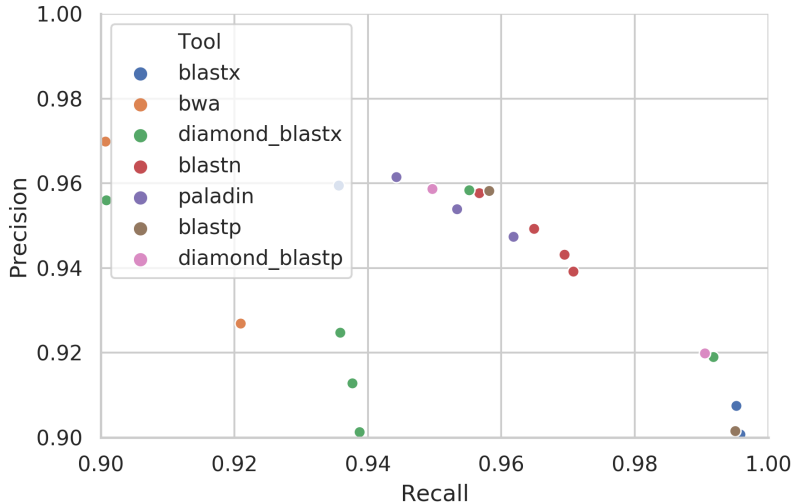
How many relevant items are selected?

$$\text{Recall} = \frac{\text{true positives}}{\text{true positives} + \text{false negatives}}$$

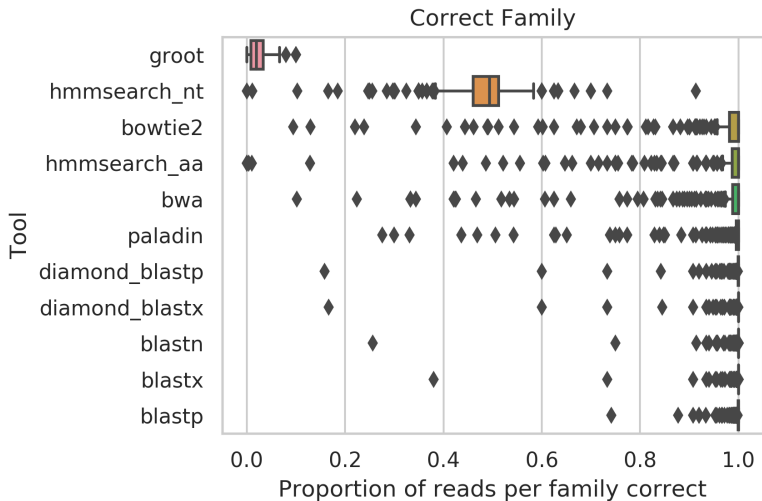
bit.ly/2pZzxJU

**How well do different methods
do?**

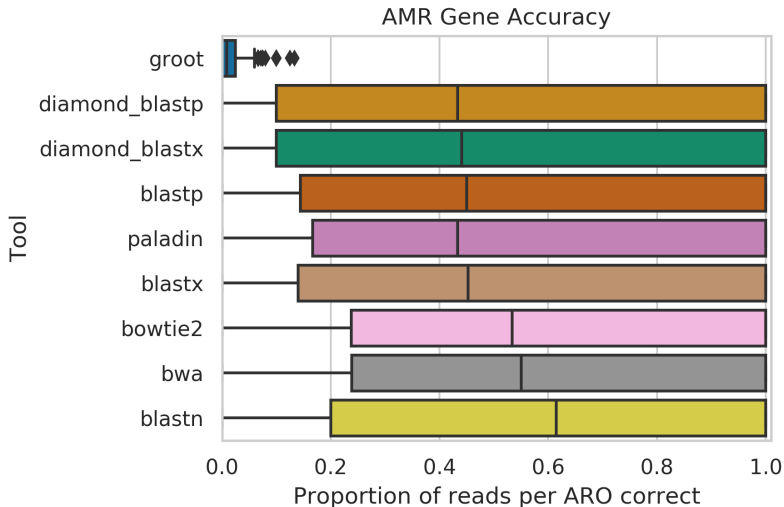
We can find reads from AMR genes



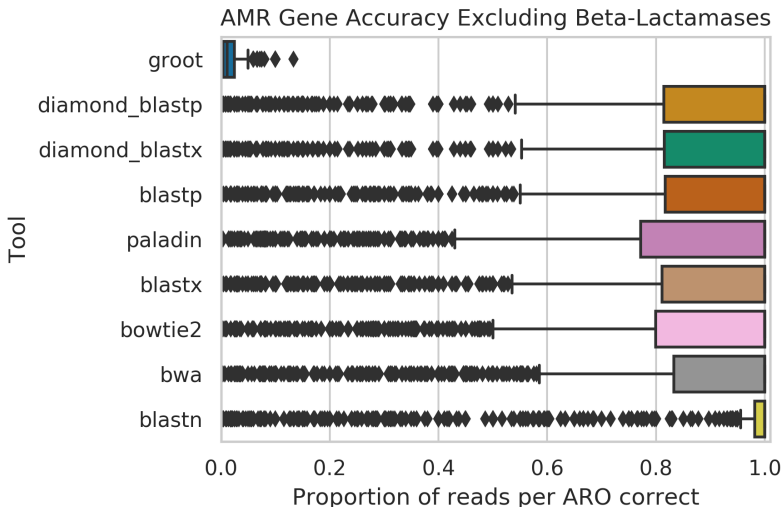
We can mostly identify which family



We cannot identify which specific gene

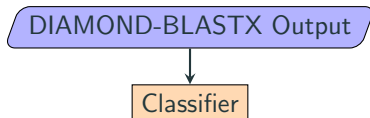


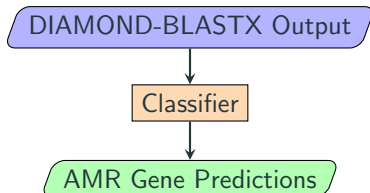
Highly similar families to blame

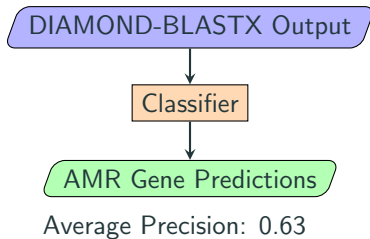


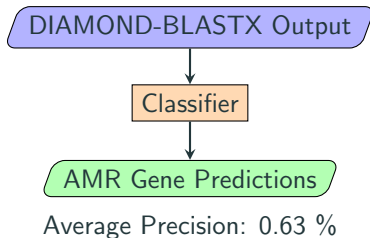
Is there any way to improve this?

DIAMOND-BLASTX Output



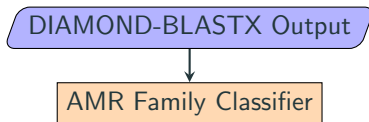




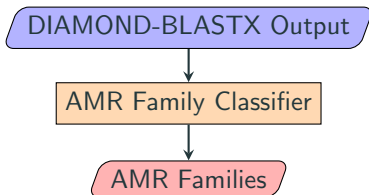


DIAMOND-BLASTX Output

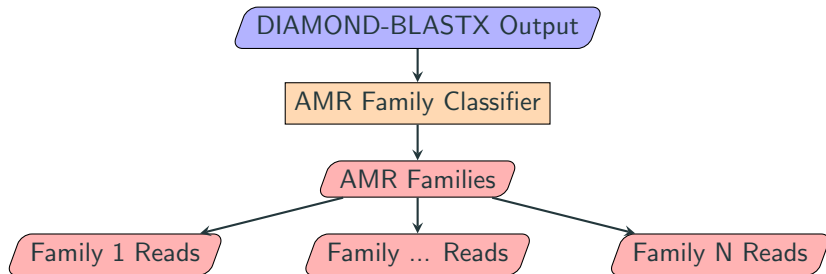
Revised classifier structure: exploiting the ARO



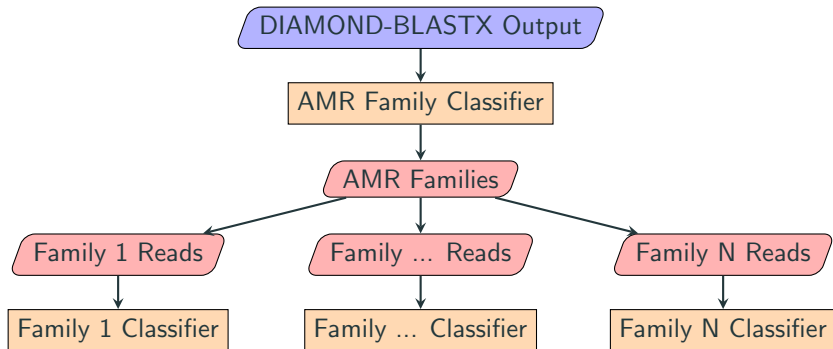
Revised classifier structure: exploiting the ARO



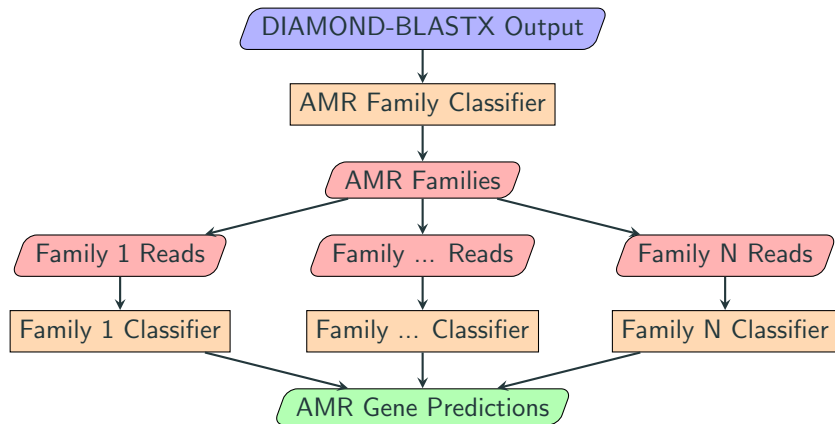
Revised classifier structure: exploiting the ARO



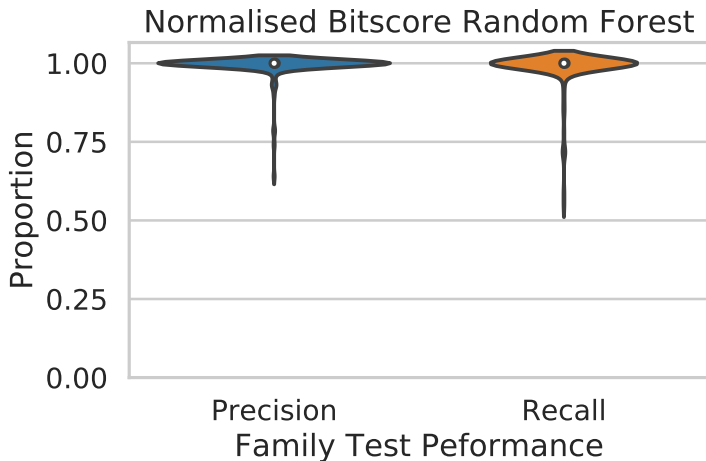
Revised classifier structure: exploiting the ARO



Revised classifier structure: exploiting the ARO

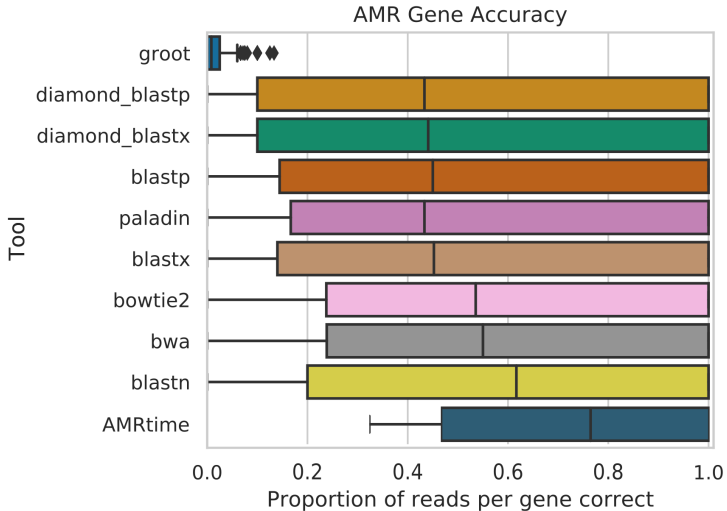


Slightly improved family performance

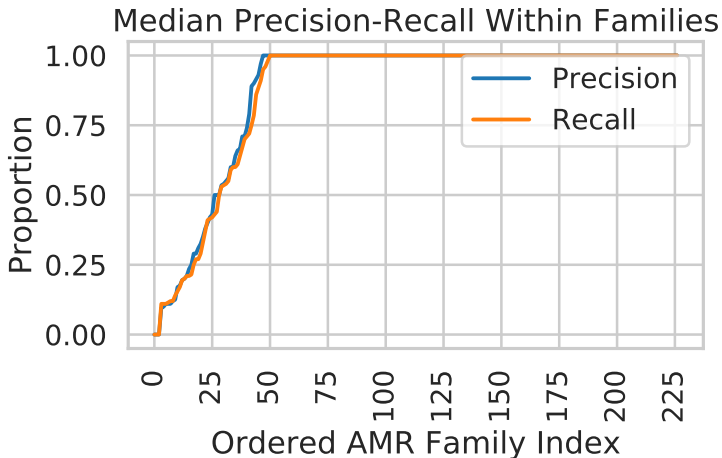


Mean Precision: 0.995, Mean Recall: 0.985

Greatly improved gene performance

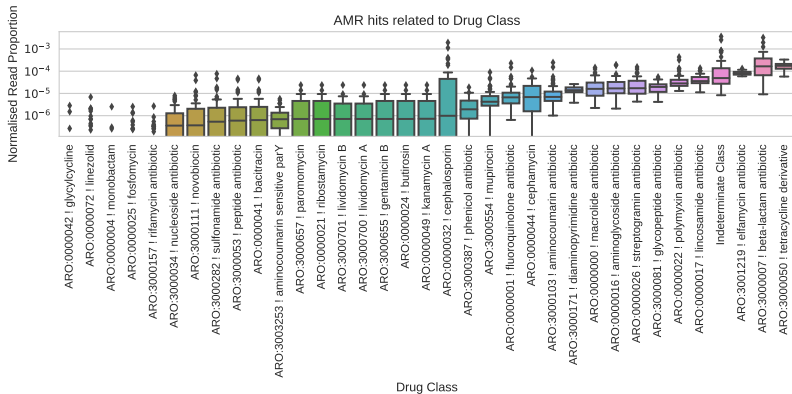


Gains not evenly distributed



- Not enough signal in read so output compatible set
- Some fixed bugs

Metagenomic resistome profile



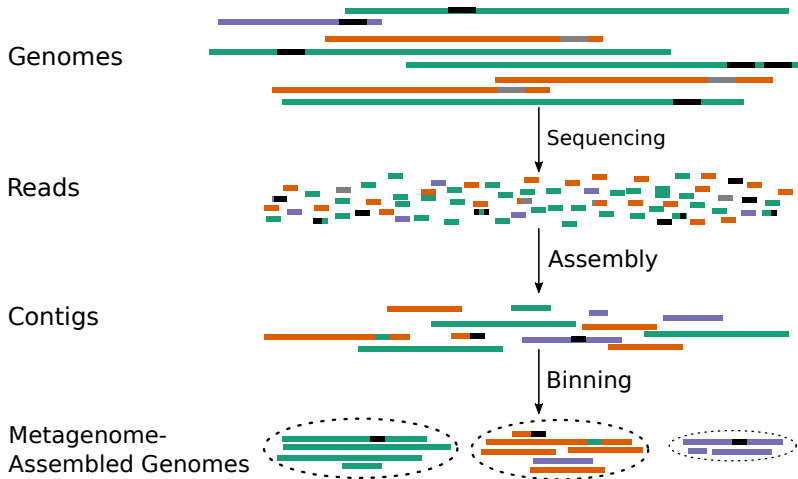
47 human gut metagenome profiles

- Known AMR genes
- Is one organism resistant to everything?
- Are many organisms each resistant to one thing?
- Have AMR genes been laterally transferred?

**Can we get the best of
metagenomics and genomics?**

Metagenomic-Assembled Genomes

MAG binning



MAGs are popular

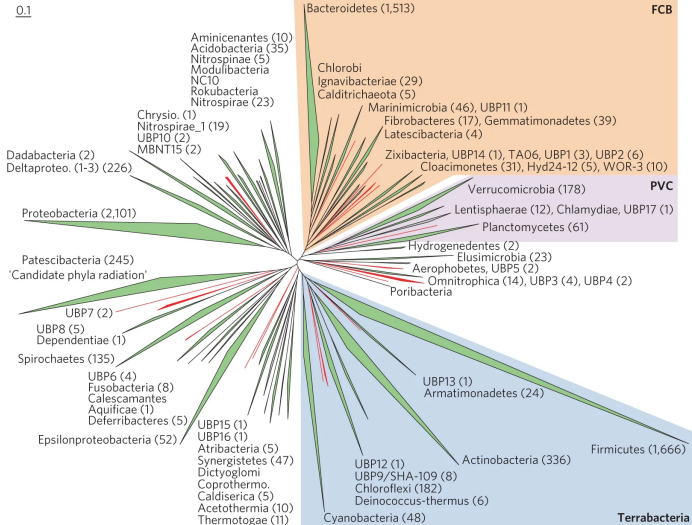


Figure from (Parks et al., 2017)

What about plasmids?

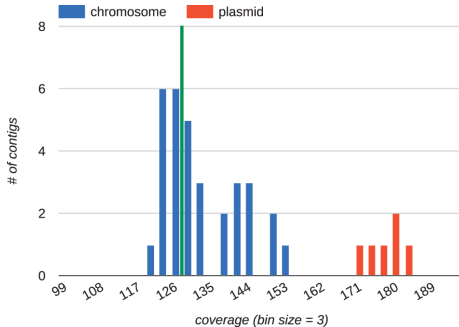
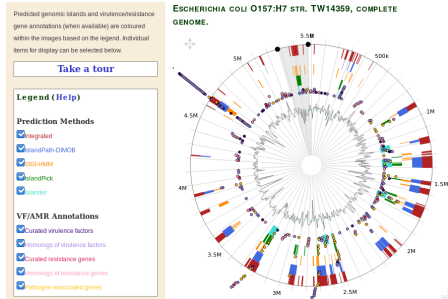


Figure from (Antipov et al., 2016)

- Circular or linear extrachromosomal self-replicating DNA.
- Dissemination of AMR genes.
- Repetitive, variable copy number, different sequence composition.

Or genomic islands



www.pathogenomics.sfu.ca/islandviewer

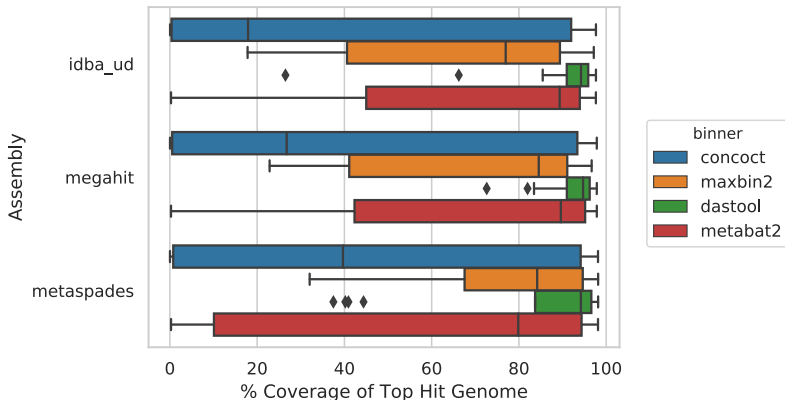
- Clusters of genes acquired through LGT
- Integrons, transposons, integrative and conjugative elements (ICEs) and prophages
- Variable copy number and composition (used by SIGI-HMM, IslandPath-DIMOB)

How well do MAGs recover these sequences?

Time to start simulating again

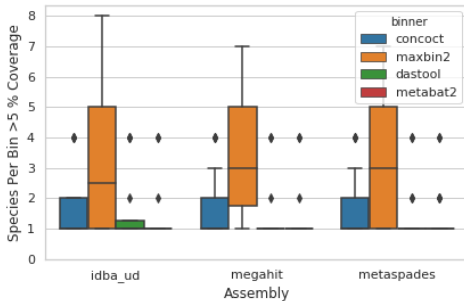
- Simulate some metagenomes (lognormal abundance distribution) from difficult genomes
 - 10 genomes: lots of plasmids
 - 10 genomes: high % of genomic islands (compositional)
 - 10 genomes: low % of genomic islands
- Assembly using 3 alternative methods: IDBA_UD, MetaSPAdes, Megahit
- Bin contigs using 4 different tools: metabat2, maxbin2, concoct, dastool

Chromosomes fairly well binned



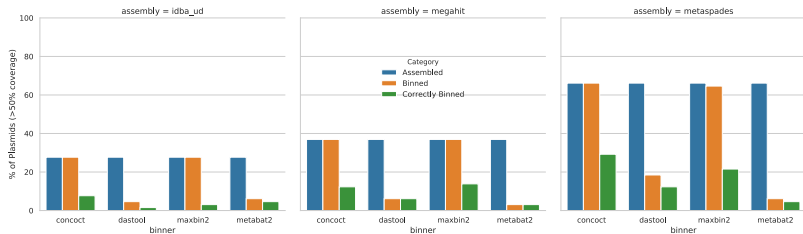
26-94.3% median chromosomal coverage (Pre-print draft
github.com/fmaguire/mag_sim_paper)

Chromosomes fairly well binned



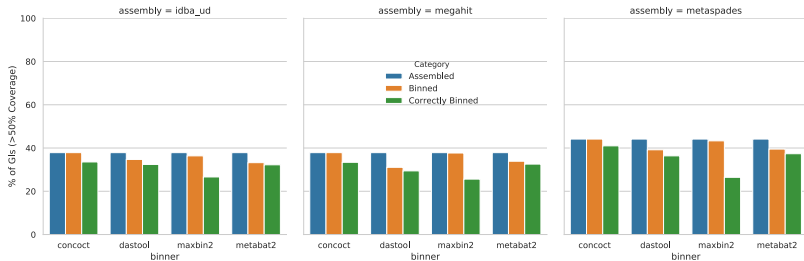
26-94.3% median chromosomal coverage (Pre-print draft
github.com/fmaguire/mag_sim_paper)

Plasmids are not



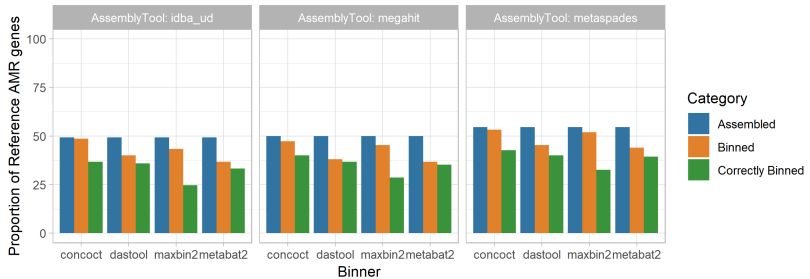
1.5-29.2% plasmids binned

Genomic islands are better but bad



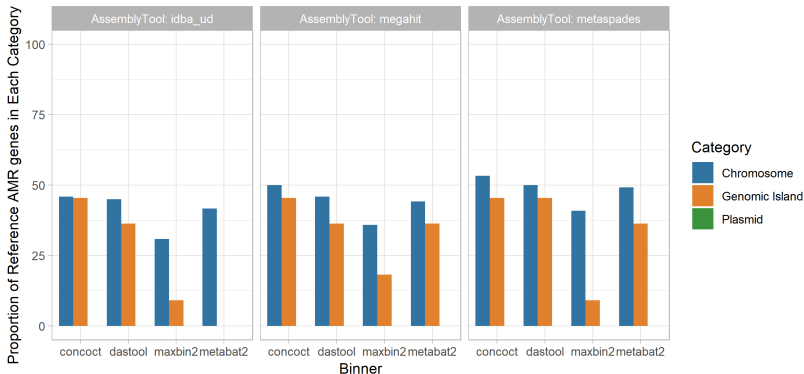
28-42% GIs binned

What about AMR genes?



24-43% AMR genes binned

Which AMR genes are lost?



- 30-53% chromosomal AMR genes (n=120)
- 0-45% genomic island AMR genes (n=11)
- 0% of plasmid AMR genes (n=20)

Be cautious with MAGs

- Regain some context but with biased data loss
- Disproportionate loss of AMR genes
- Mobile Genetic Elements poorly recovered
- Cautionary tale: more processing = more data loss

Conclusions

Conclusions

Method	Strengths	Weaknesses
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Conclusions

Method	Strengths	Weaknesses
Targeted	Cheap, easy analysis	<i>a priori</i> , stagnation

Conclusions

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Targeted	Cheap, easy analysis	<i>a priori</i> , stagnation
Genomics	Context, moderate analysis	Isolation, throughput

Conclusions

Method	Strengths	Weaknesses
Targeted	Cheap, easy analysis	<i>a priori</i> , stagnation
Genomics	Context, moderate analysis	Isolation, throughput
Metagenomics	Many genomes at once	Fragmented, no context, difficult analysis

Conclusions

Method	Strengths	Weaknesses
Targeted	Cheap, easy analysis	<i>a priori</i> , stagnation
Genomics	Context, moderate analysis	Isolation, throughput
Metagenomics	Many genomes at once	Fragmented, no context, difficult analysis
Metagenomic-Assembled Genomes	Context for many genomes	Lose key data, complex analysis

- Simulation fundamental to evaluating approaches

Conclusions

Method	Strengths	Weaknesses
Targeted	Cheap, easy analysis	<i>a priori</i> , stagnation
Genomics	Context, moderate analysis	Isolation, throughput
Metagenomics	Many genomes at once	Fragmented, no context, difficult analysis
Metagenomic-Assembled Genomes	Context for many genomes	Lose key data, complex analysis

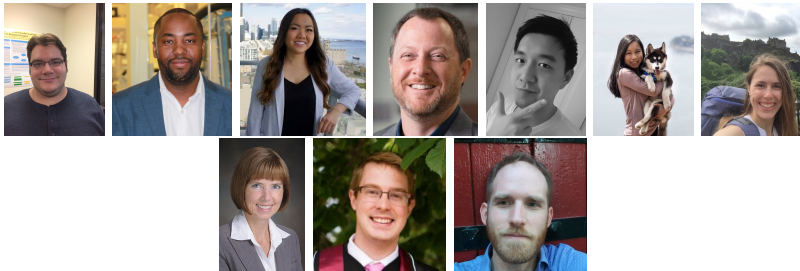
- Simulation fundamental to evaluating approaches
- Characterisation necessary to mitigate weaknesses and promote strengths

Conclusions

Method	Strengths	Weaknesses
Targeted	Cheap, easy analysis	<i>a priori</i> , stagnation
Genomics	Context, moderate analysis	Isolation, throughput
Metagenomics	Many genomes at once	Fragmented, no context, difficult analysis
Metagenomic-Assembled Genomes	Context for many genomes	Lose key data, complex analysis

- Simulation fundamental to evaluating approaches
- Characterisation necessary to mitigate weaknesses and promote strengths
- Machine-Learning represents useful tools for this (e.g. AMRtime, gene-free AST prediction models)

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Questions?

References

- Antipov, D., Hartwick, N., Shen, M., Raiko, M., Lapidus, A., and Pevzner, P. (2016). plasmidspades: assembling plasmids from whole genome sequencing data. *bioRxiv*, page 048942.
- Bradley, P., Gordon, N. C., Walker, T. M., Dunn, L., Heys, S., Huang, B., Earle, S., Pankhurst, L. J., Anson, L., De Cesare, M., et al. (2015). Rapid antibiotic-resistance predictions from genome sequence data for staphylococcus aureus and mycobacterium tuberculosis. *Nature communications*, 6:10063.

- Cassini, A., Hogberg, L. D., Plachouras, D., Quattrocchi, A., Hoxha, A., Simonsen, G. S., Colomb-Cotin, M., Kretzschmar, M. E., Devleesschauwer, B., Cecchini, M., et al. (2019). Attributable deaths and disability-adjusted life-years caused by infections with antibiotic-resistant bacteria in the eu and the european economic area in 2015: a population-level modelling analysis. *The Lancet Infectious Diseases*, 19(1):56–66.
- de Kraker, M. E., Stewardson, A. J., and Harbarth, S. (2016). Will 10 million people die a year due to antimicrobial resistance by 2050? *PLoS medicine*, 13(11):e1002184.
- Goossens, H., Ferech, M., Vander Stichele, R., Elseviers, M., Group, E. P., et al. (2005). Outpatient antibiotic use in europe and association with resistance: a cross-national database study. *The Lancet*, 365(9459):579–587.

- Guiton, A. K., Raphenya, A. R., Klunk, J., Kuch, M., Alcock, B., Surette, M. G., McArthur, A. G., Poinar, H. N., and Wright, G. D. (2019). Capturing the resistome: A targeted capture method to reveal antibiotic resistance determinants in metagenomes. *Antimicrobial agents and chemotherapy*, pages AAC-01324.
- Maguire, F. (2016). A multi-omic analysis of the photosynthetic endosymbioses of paramecium bursaria. *PhD Thesis*.
- Maguire, F., Rehman, M. A., Carrillo, C., Diarra, M. S., and Beiko, R. G. (2019). Identification of primary antimicrobial resistance drivers in agricultural nontyphoidal salmonella enterica serovars by using machine learning. *MSystems*, 4(4):e00211-19.
- Matthews, T. C., Bristow, F. R., Griffiths, E. J., Petkau, A., Adam, J., Dooley, D., Kruczkiewicz, P., Curatcha, J., Cabral, J., Fornika, D., et al. (2018). The integrated rapid infectious disease analysis (irida) platform. *bioRxiv*, page 381830.

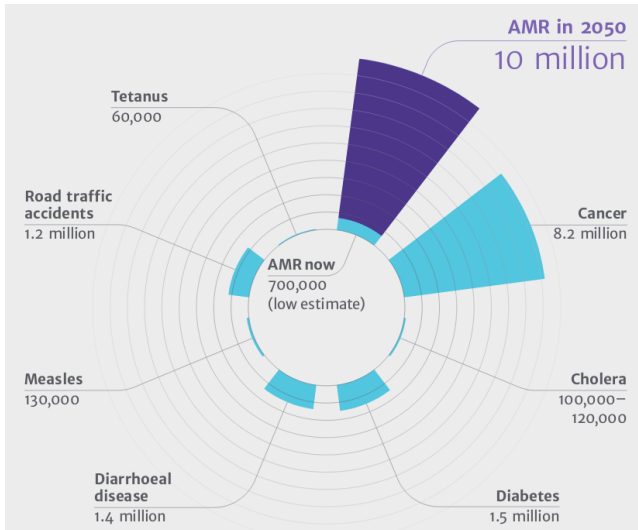
on Antimicrobial Resistance, R. (2016). *Tackling drug-resistant infections globally: final report and recommendations*. Review on antimicrobial resistance.

Parks, D. H., Rinke, C., Chuvochina, M., Chaumeil, P.-A., Woodcroft, B. J., Evans, P. N., Hugenholtz, P., and Tyson, G. W. (2017). Recovery of nearly 8,000 metagenome-assembled genomes substantially expands the tree of life. *Nature microbiology*, 2(11):1533.

Stairs, J., Bal, N., Maguire, F., and Scott, H. (2019). A resident-led clinic that promotes the health of refugee women through advocacy and partnership. *Canadian Medical Education Journal*.

Backup

10 million deaths?



(on Antimicrobial Resistance, 2016), (de Kraker et al., 2016)

10 million deaths?

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PMCID: PMC5127510

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PMID: [27898664](https://pubmed.ncbi.nlm.nih.gov/27898664/)

Will 10 Million People Die a Year due to Antimicrobial Resistance by 2050?

[Marieke E. A. de Kraker](#),^{1,*} [Andrew J. Stewardson](#),² and [Stephan Harbarth](#)¹

(on Antimicrobial Resistance, 2016), (de Kraker et al., 2016)

Where does 10 million come from?

For 3rd-generation cephalosporin resistant *E. coli*, *K. pneumoniae*, and MRSA:

- Estimate global BSIs (multiply average incidence in tertiary European hospitals by global population).
- Estimate AMR (proportion of resistant blood-cultures per country)
- Extrapolate to other infections sites (via relative incidence to BSI in 2 studies n=16 BSIs)
- Estimate attributable mortality rates from adjusted odds-ratios in an unspecified manner.
- Assume no change in mortality, 40% increase in resistance, and doubled infection rates by 2050.