

RNAi pathway components and function in *Paramecium bursaria*

Finlay Maguire

University of Exeter

June 2, 2016

Overview

Motivation

RNAi in ciliates

Experimental RNAi induction in *P. bursaria*

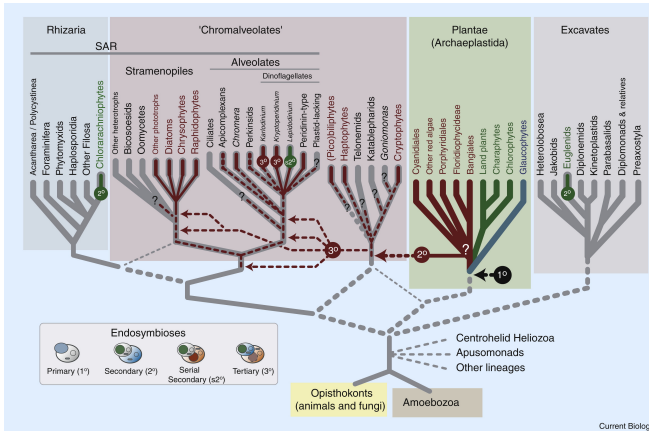
RNAi pathway components in active *P. bursaria* transcriptome(s)

In-silico analysis of potential endosymbiont 'cross-talk'

Conclusions

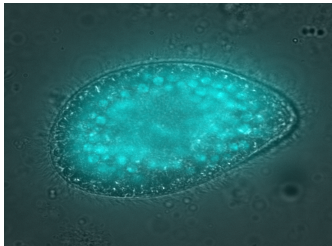
Why is *Paramecium bursaria*
potentially a good model for
(secondary photosynthetic)
endosymbiosis?

Broad diversity of plastid endosymbioses



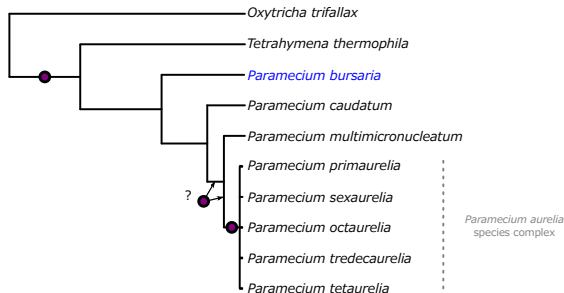
Reproduced from [Arc09].

Paramecium bursaria and its green algal endosymbionts



- ▶ 100 μm to 160 μm serial phagotrophic ciliate (nuclear dimorphism).
- ▶ ~ 300 endosymbiotic algae in stable heritable facultative(?) endosymbiosis.
- ▶ Multiple independent origins of these endosymbioses.
- ▶ Single cell transcriptome and genome of *P. bursaria*-*Micractinium reisseri* CCAP 1660/12.
- ▶ *P. bursaria* bulk transcriptome Yad1g1N [KSD⁺14].

RNAi pathways in the ciliates



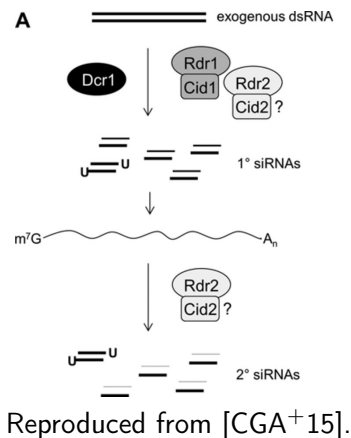
A good model needs a means to test hypotheses:

- ▶ Ciliate specific scnRNA system [MG04, CMM13].
- ▶ siRNA pathways present in *Paramecium tetaurelia* [GS01, GS02] (and *Tetrahymena thermophila* [CL06, YC05]):
 1. Transgene inducible pathway [GS01].
 2. Exogenous dsRNA inducible pathway (feeding or injection) [GS02].

Transgene pathway

- ▶ Microinjection and transformation of MAC with high-copy transgenes lacking 3' UTR [GS01]:
 1. 23nt siRNA generated from transgene transcripts (Dcr1, Rdr2, Rdr3 and Cid2) [LNS⁺09, MCT⁺14].
 2. mRNA cleavage (Ptiwi13 and Ptiwi14).

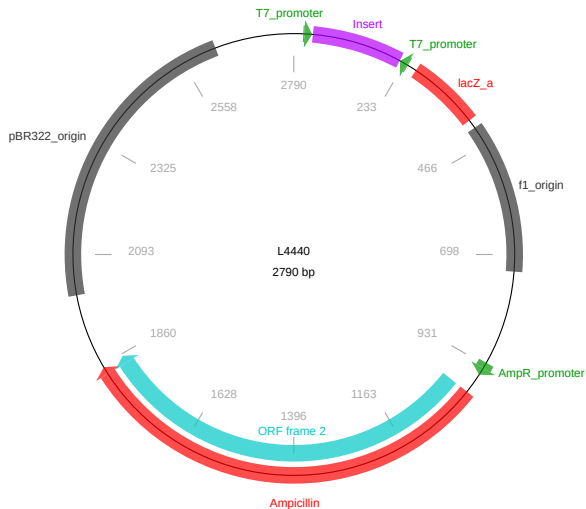
Exogenous dsRNA pathway



- ▶ Exogenous dsRNA via feeding (or microinjection) [GS02].
- ▶ 1° siRNA targeted cleavage (Ptiwi13) [BGK⁺11].
- ▶ Undefined role in MAC for 2° siRNA (Ptiwi12, Ptiwi15) [MCT⁺14, CGA⁺15, BGK⁺11].
- ▶ Pds1 involved in uptake of dsRNA from vacuole? [CGA⁺15].
- ▶ Activated at low levels by ssRNA from normal food bacteria [CGA⁺15].

So, can we experimentally induce
RNAi in *P. bursaria*?

Experimental feeding vector



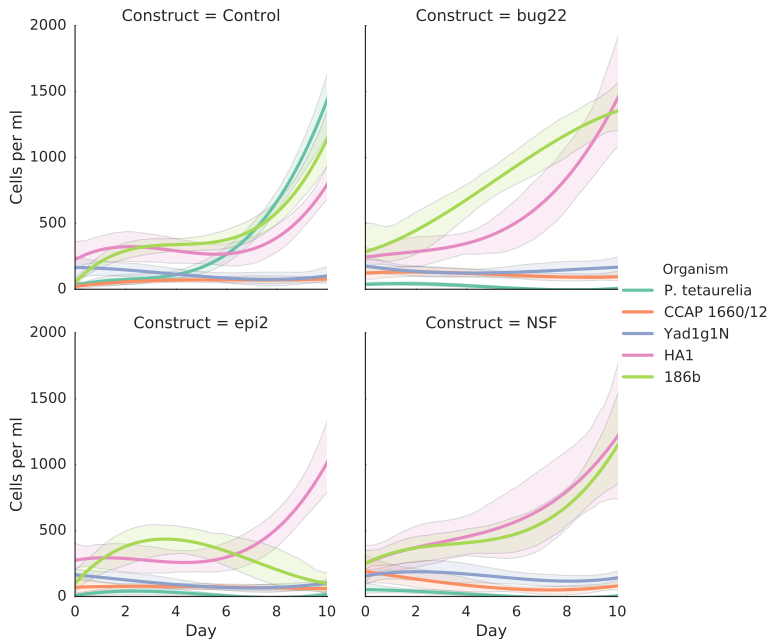
Transformed into *E. coli* with IPTG-inducible T7 polymerase and RNase III deficiency.

Construct inserts

Gene	Function	RNAi phenotype in <i>P. tetraurelia</i>
<i>epi2</i>	Epiplasmin	"Monstrous" cells
NSF	Membrane fusion factor	Lethal
<i>bug22</i>	Basal body/ciliary protein	Slow swimming and death



RNAi feeding had mixed results



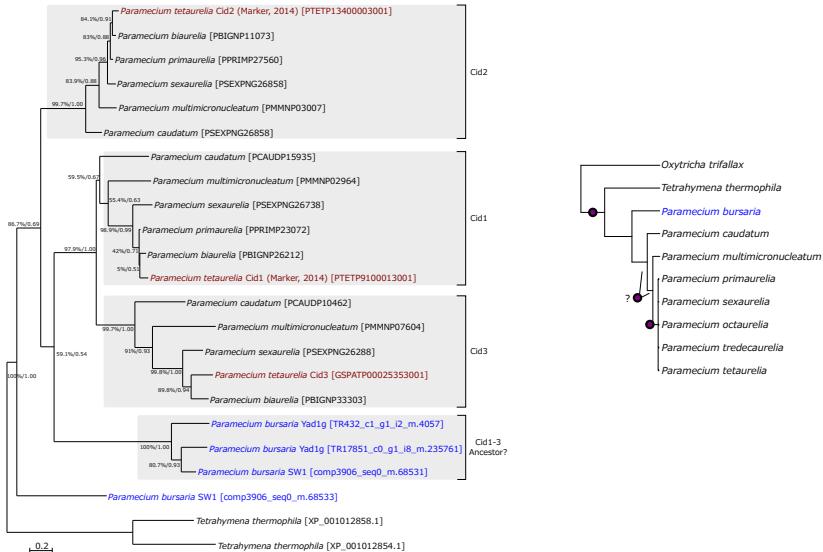
Are all the known RNAi pathway components present in the active transcriptome(s)?

Summary of known RNAi components

Pathway	Component	Function
transgene-induced siRNA	Rdr3 Ptiwi14	RdRP Piwi
both pathways	Rdr2 Dcr1 Ptiwi13 Cid2	RdRP Dicer Piwi Nucleotidyl transferase
exogenous dsRNA-induced siRNA	Rdr1 Cid1 Ptiwi12 Ptiwi15 Pds1	RdRP Nucleotidyl transferase Piwi Piwi Import of dsRNA?

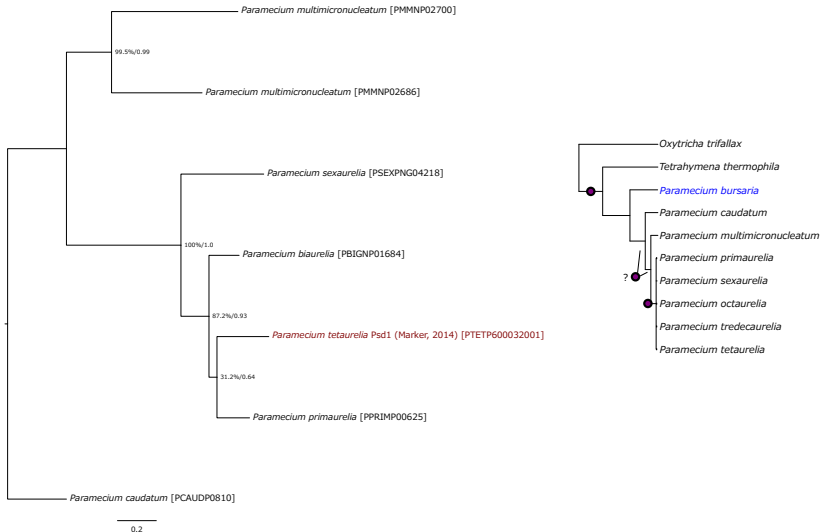
Cid ancestor

Cid

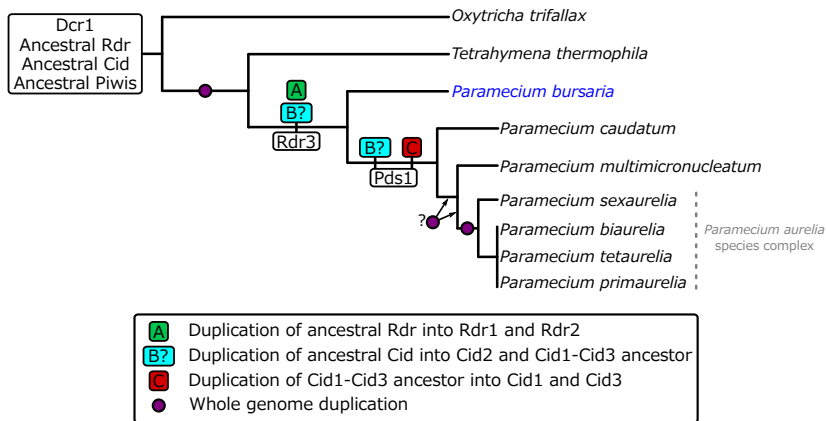


Pds1 absent

Pds1

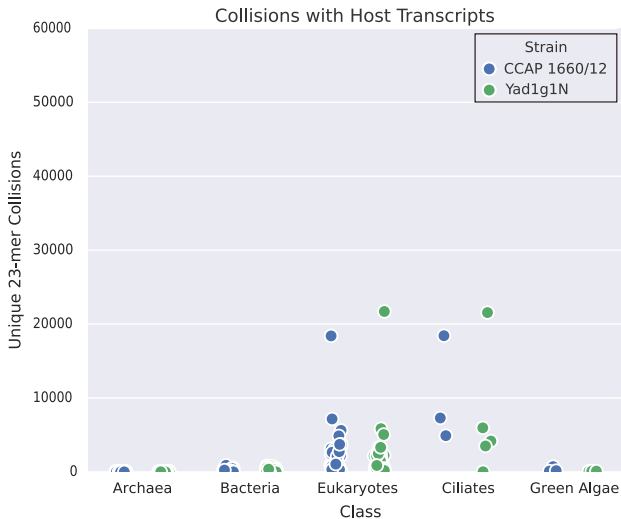


Putative RNAi component evolution scenario

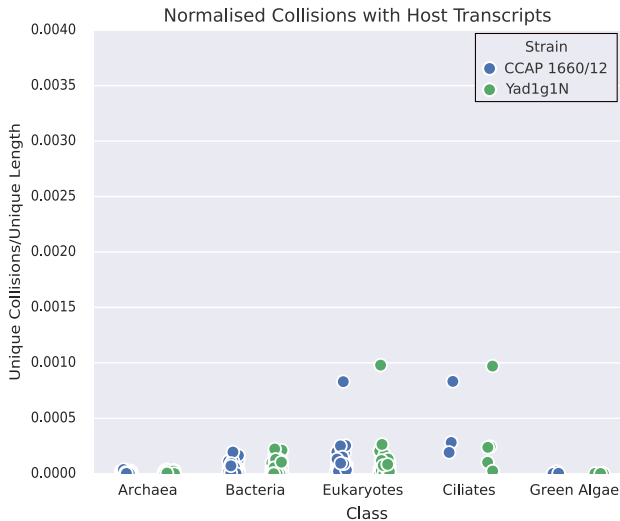


Could having a eukaryotic endosymbiont and RNAi activated by dsRNA in vacuoles be deleterious?

Higher level of collisions with eukaryotes



Collisions are a function of transcriptome size



Conclusions

- ▶ RNAi phenotypes not inducible in most *P. bursaria* strains via feeding.
- ▶ *P. bursaria* lacks Pds1 (in active transcriptome) thus may be unable to take up RNA from digestive vacuoles.
- ▶ High levels of 23-mer collisions between *P. bursaria* and eukaryotic endosymbiont transcriptomes may lead to deactivation of dsRNA uptake from vacuoles.
- ▶ Presence of other factors in active transcriptomes of *P. bursaria* indicate transgene and microinjected exogenous dsRNA pathways may function.

Acknowledgements

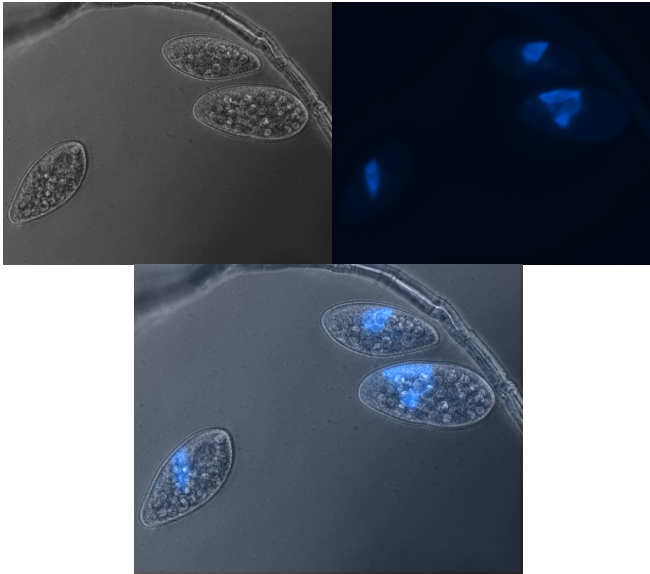
- ▶ Ben Jenkins (feeding experiments)
- ▶ David Milner (labwork)
- ▶ Tom Richards (PI)
- ▶ NHM-UCL PhD Studentship (main funding)



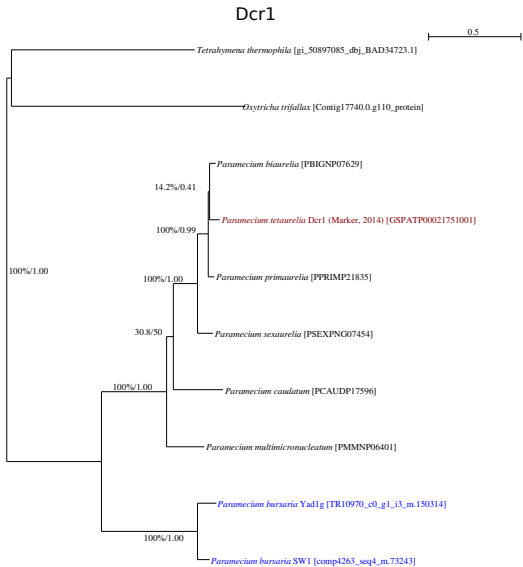
References

- [Arc09] John M Archibald. The puzzle of plastid evolution. *Curr. Biol.*, 19(2):R81–8, January 2009.
- [BGK⁺11] K. Bouhouche, J.-F. Gout, a. Kapusta, M. Betermier, and E. Meyer. Functional specialization of Piwi proteins in *Paramecium tetraurelia* from post-transcriptional gene silencing to genome remodelling. *Nucleic Acids Res.*, 39(10):4249–4264, 2011.
- [CGA⁺15] Q. Carradec, U. Gotz, O. Arnaiz, J. Pouch, M. Simon, E. Meyer, and S. Marker. Primary and secondary siRNA synthesis triggered by RNAs from food bacteria in the ciliate *Paramecium tetraurelia*. *Nucleic Acids Res.*, 43(3):1818–1833, 2015.
- [CL06] Kathleen Collins and Suzanne R Lee. Two classes of endogenous small RNAs in *Tetrahymena thermophila*. *Genes Dev.*, 20:28–33, 2006.
- [CMM13] Douglas L Chalker, E. Meyer, and Kazufumi Mochizuki. Epigenetics of ciliates. *Cold Spring Harb. Perspectives Epigenetics*, 5:a017764, 2013.
- [GS01] Angélique Galvani and Linda Sperling. Transgene-mediated post-transcriptional gene silencing is inhibited by 3 non-coding sequences in *Paramecium*. *Nucleic Acids Res.*, 29(21):4387–4394, 2001.
- [GS02] Angélique Galvani and Linda Sperling. RNA interference by feeding in *Paramecium*. *Trends Genet.*, 18(1):11–2, January 2002.
- [KSD⁺14] Yuuki Kodama, Haruo Suzuki, Hideo Dohra, Manabu Sugii, Tatsuya Kitazume, Katsushi Yamaguchi, Shuji Shigenobu, and Masahiro Fujishima. Comparison of gene expression of *Paramecium bursaria* with and without *Chlorella variabilis* symbionts. *BMC Genomics*, 15:183, 2014.
- [LNS⁺09] G. Lepere, M. Nowacki, V. Serrano, J.-F. Gout, G. Guglielmi, S. Duhaucourt, and E. Meyer. Silencing-associated and meiosis-specific small RNA pathways in *Paramecium tetraurelia*. *Nucleic Acids Res.*, 37(3):903–915, 2009.
- [MCT⁺14] Simone Marker, Quentin Carradec, Véronique Tanty, Olivier Arnaiz, and Eric Meyer. A forward genetic screen reveals essential and non-essential RNAi factors in *Paramecium tetraurelia*. *Nucleic Acids Res.*, 42(11):7268–7280, 2014.
- [MG04] Kazufumi Mochizuki and Martin a. Gorovsky. Conjugation-specific small RNAs in *Tetrahymena* have predicted properties of scan (scn) RNAs involved in genome rearrangement. *Genes Dev.*, 18(Nanney 1974):2068–2073, 2004.
- [YC05] Meng-Chao Yao and Ju-Lan Chao. RNA-guided DNA deletion in *Tetrahymena*: an RNAi-based mechanism for programmed genome rearrangements. *Annu. Rev. Genet.*, 39:537–59, 2005.

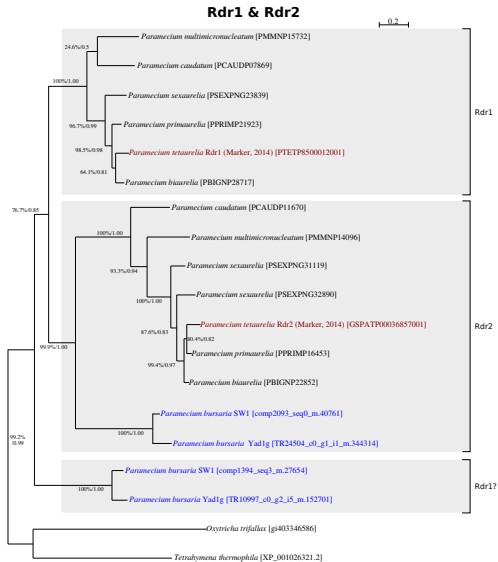
Microinjection proved difficult



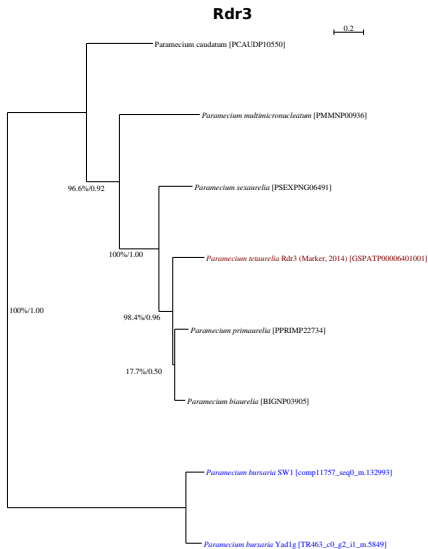
Dcr1



Rdr1 Rdr2



Rdr3



Psd1 Structure

